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Instruction For Use

SirYus CoViDiag+®

Research Use Only device for the anti-SARS-CoV-2&variants antibodies detection and identification

For Research Use Only

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1. INTENDED USE

The SirYus CoViDiag kit is intended for the qualitative determination of IgG class antibodies against SARS-CoV-2 and some of the variants in human serum or plasma to support the diagnosis of COVID-19 infection.

This kit constitutes a supplement to direct pathogen detection.

In addition, serology can be used to collect epidemiological information on the prevalence of SARS-CoV-2.

2. CONTEXT

End of 2019, a novel respiratory disease emerged in the city of Wuhan, Hubei Province of the People's Republic of China, and soon spread rapidly within the country and worldwide. The causative agent was identified as severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2).

The clinical manifestations of SARS-CoV-2-related COVID-19 infection include fever, cough, respiratory problems and fatigue. In most patients the affection shows symptoms of a mild febrile illness with irregular lung infiltrates.

Different methodologies of diagnostic tests are available:

- Molecular tests for the detection of the SARS-CoV-2 coronavirus genome by RT-PCR which is, at present, the only technique for COVID-19 diagnostic recommended,
- Antigenic tests which allow the detection of specific SARS-CoV-2 proteins. Given their poor performance, these antigenic tests are not currently recommended,
- Serological tests that intent to detect specific antibodies (Ab) produced by the organism and directed against SARS-CoV-2.

Few consolidated data are currently available on the immune response against SARS-CoV-2. However, based on the first published studies, a set of observations agree on several aspects of the adaptive humoral immune response, and more particularly on the kinetics of anti-SARS-CoV-2 antibody production.

The IgG production is detectable in symptomatic patients from the second week after onset of symptoms. However, cases have also been reported with later antibody production, beyond the 15th day after the onset of symptoms, and up to 30 days after infection, especially in asymptomatic or paucisymptomatic patients.

Serological tests cannot tell whether the person is contagious or not. This is because seroconversion is not accompanied by a drop in viral titer. There is no established correlation between the production of antibodies and the presence of the infectious virus. In addition, neither the infectious dose of SARS-CoV-2 nor the dose initiating an antibody response is currently known.

Serologic testing can only determine whether a person has produced antibodies in response to infection with the virus, in other words, whether or not that person has triggered an immune response against the virus. But an immune response is not always a proof of protective immunization against further infection with the same virus. To get a protective immunization, it is necessary that the organism produces high titers of antibodies that prevents the action of the virus and in particular its entry into the target cells. We then speak about neutralizing antibodies.

In addition, these important titers of neutralizing antibodies have to be produced over a long period to ensure a protection over the time. However, the target epitopes of neutralizing antibodies have not yet been identified.

Regarding the viral targets used in serological tests to detect IgG, two proteins are frequently used: protein S (spike), the surface protein of the virus allowing interaction (via its RBD domain) and fusion with the target cell and the N protein (core protein, internal to the virus). While the S protein (or its RBD domain) has been described as inducing an earlier response or better correlated with the presence of neutralizing antibodies, the precise neutralizing epitopes have not been characterized. These epitopes may also depend on the conformation of the protein, in particular as regards the S protein. To date, there is therefore no robust element to promote the use of a precise form or of another of these proteins.

Serological tests are performed on blood samples and are intended to identify patients who have developed immunity to SARS-CoV-2 whether they have been symptomatic or not. Serological tests could identify in certain circumstances the patients being or having been infected by SARS-CoV-2 or monitor the serological status of exposed persons (healthcare professionals for example). Finally, these tests can also be useful in collecting epidemiological data related to COVID-19 (actually infected patients, rate of severe forms, etc.) or open perspectives for prognostic data.

3. PRINCIPLE OF THE TEST

The qualitative immunoenzymatic detection of specific antibodies is based on ELISA technology principle. Regarding SirYus CoViDiag kit, 7 different ELISA-like assays are miniaturized and combined.

Microtitration plates wells were printed with 7 different antigens from SARS-CoV-2 separately. They are dedicated to bind specific antibodies in the tested samples and therefore deliver 7 different responses in one single assay.

Printed antigens are :

- N protein (Nucleocapside protein, internal to the virus),
- S1 protein (Sub-unit 1 of the Spike protein),
- RBD wt (Receptor Binding Domain) of Spike 1 protein, wt
- RBD B.1.1.7 (Receptor Binding Domain) of Spike 1 protein, strain B.1.1.7 (UK variant),
- RBD B.1.351 (Receptor Binding Domain) of Spike 1 protein, strain B.1.351 (SA variant),
- NTD domain of the S1 protein (N-Terminal Domain),
- S2 protein (Sub-unit 2 of the Spike protein).

Diluted sample (sera or plasma) is incubated in a microtitration plate well. After a washing step to remove the sample, a secondary antibody conjugated to an enzyme (HRP) is added. This conjugate binds to antibodies already captured from the sample. A second washing step is dedicated to remove excess of unbound conjugate. A chromogen substrate: tétraméthylbenzidine (TMB), is then used in order to detect the antibody's complexes. Distinguishable individual circular blue precipitates will be visible at the bottom of the well when such complexes are present.

The color intensity depends on the amount of specific antibodies present in the sample.

For results interpretation, the well bottom surfaces can be analysed with bare-eyes or imaged using a colorimetric reader.

4. MATERIAL

4.1 Reagents

SC | CVD **Precoated plate: 12 breakable strips of 8 wells in a microplate** ; each well were printed with 5 different antigens of SARS-CoV-2 ; plates are packed in an aluminium sealed foil.

DB | CVD **Dilution Buffer** : 1 x 120 mL bottle of aqueous buffer for sample dilution ; Ready to use ; orange color; with <0.05% Proclin® 300.

WB | CVD **Wash Buffer** : 2 x 120 mL bottles of aqueous buffer for wells washing; ready to use ; blue color ; with <0.05% Proclin® 300.

CA | CVD **Conjugate Ab** : 1 x 100 µL vial of anti-IgG human conjugate to a HRP peroxydase; to be diluted ; Blue cap ;with <0.05% Proclin® 300.

SU | CVD **Substrate** : 1 x 15 mL bottle of insoluble chromogen substrate (3,3',5,5'-Tetramethylbenzidine) for HRP enzyme; ready to use ; brown bottle.

An instruction for use booklet is provided with the kit.

For hazard and precautionary statements see chapter 10.2

For potential hazardous substances please check the safety data sheet.

4.2 Materials and equipment required

- Incubator 37°C
- Deionised water
- Pipettes to deliver volumes between 10 and 1000 µL
- Vortex tube mixer
- 96 wells microplate plate sticker or cover foil
- Disposable tubes or polypropylene plate

4.3 Optional materials and equipment

- Automatic microtiter plate washer
- Microtiter plate Agitator
- Multi-channels pipette
- Colorimetric reader for biochips

5. STABILITY AND STORAGE

Store the kit at 2...8 °C.

Expiry date is indicated on each composant of the kit.

Expiry date of the kit correspond to the expiry date of the composant with the shortest shelf life.

Performances of the test are depending on expiry date. Do not use reagents and tests after the expiry date.

Sample and conjugate dilutions have to be performed for each assay.

6. REAGENTS PREPARATION

It is very important to bring all reagents and samples to room temperature (20...25 °C) 30 min prior to the test and mix them before starting the test run!

6.1 Test well strips

The wells of the plate/breakable strips were printed with 5 different antigens from SARS-CoV-2 (fig1). Immediately after removal of the strips, the remaining strips should be resealed in the aluminium foil along with the desiccant supplied.

Remaining non-use strips in sealed foil have to stored at 2...8 °C.

| | | | | | | | | | | |
|----|------------------|-------|----------------|----------------|----------------|------|------|------|-------|-------|
| | 1 | 2 | 3 | 4 | 5 | 6 | 7 | 8 | 9 | 10 |
| 1 | CTRL+ | CTRL+ | | | | | | | CTRL+ | CTRL+ |
| 2 | | | RBD B.1.1.7 | RBD B.1.1.7 | RBD B.1.1.7 | | NTD | NTD | NTD | |
| 3 | RBD B.1.351 | | | | | | | | | |
| 4 | RBD B.1.351 | | S1 | S1 | S1 | | N | | | N |
| 5 | RBD B.1.351 | | S1 | | | | N | N | | N |
| 6 | | | S1 | S1 | S1 | | N | | N | N |
| 7 | RBD B.1.1.248 | | | | S1 | | N | | | N |
| 8 | RBD B.1.1.248 | | S1 | S1 | S1 | | N | | | N |
| 9 | RBD B.1.1.248 | | | | | | | | | |
| 10 | | | RBD wt | RBD wt | RBD wt | | S2 | S2 | S2 | |
| 11 | CTRL+ | CTRL+ | | Ctrl | Ctrl | Ctrl | Ctrl | Ctrl | | CTRL+ |

| | |
|---------------|--|
| CTRL+ | Positive control |
| SampCtrl | Sample control |
| N | (N) Nucleocapside protein |
| S1 | (S1) Sub-unit 1 of Spike protein |
| RBD wt | (RBD wt) Receptor Binding Domain of Spike 1 protein, wt |
| RBD B.1.1.7 | (RBD B.1.1.7) Receptor Binding Domain of Spike 1 protein, strain B.1.1.7, N501Y variant (UK variant) |
| RBD B.1.351 | (RBD B.1.351) Receptor Binding Domain of Spike 1 protein, strain B.1.351, N501Y.V2 variant (South Africa variant) |
| RBD B.1.1.248 | (RBD B.1.1.248) Receptor Binding Domain of Spike 1 protein, strain B.1.1.248, N501Y - K417T - E484K variant (Brazilian/Japanese variant) |
| NTD | (NTD) N-Terminal Domain of Spike 1 protein |
| S2 | (S2) Sub-unit 2 of Spike protein |

Figure 1 – Antigens layout of each wells of the SARS-CoV-2&Variants IgG Antibody Multiplex ELISA microplate

6.2 Washing buffer

The Wash Buffer **WB | CVD** is stable at 2-8 °C. In case salt crystals appear in the bottle, warm up the solution to 37 °C e.g. in a water bath. Mix well before use.

The Kit includes 2 x 120 mL of Wash Buffer **WB | CVD**. Current procedure requires 120 mL. In case of automatic washer use, do not use more than 120 mL to prime the system.

6.3 Conjugate

Conjugate has to be diluted to 6:1000 in the Dilution Buffer **DB | CVD**.

For a whole plate, we recommend to prepare 10 mL of conjugate diluted solution.

To prepare 10 mL of conjugate diluted solution, add 60 µL of Conjugate Antibody **CA | CVD** to 10 mL of Dilution Buffer **DB | CVD**.

6.4 Substrate solution

The solution **SU | CVD** is ready to use and has to be stored at 2-8 °C protected from the light. The solution should be colourless or could have a slight blue tinge. If the substrate turns into blue, it may have been contaminated and should be thrown away.

7. SAMPLES COLLECTION AND PREPARATION

7.1 Samples

Use human serum or plasma (citrate, heparin) samples with this assay. If the assay is performed within 5 days after sample collection, the samples should be kept at 2...8 °C; otherwise they should be aliquoted and stored deep-frozen (-70...-20 °C). If samples are stored frozen, mix thawed samples before testing. Avoid repeated freezing and thawing. Heat inactivation of samples is not recommended.

7.2 Sample dilution

All samples should be diluted 1+100 with Dilution Buffer **DB | CVD**. We recommend to dispense 10 µL sample and 1 mL Dilution Buffer **DB | CVD** into tubes or polypropylene plate to obtain a 1+100 dilution and thoroughly mix before incubation.

8. PROCEDURE OF THE TEST

Please read the instruction for use carefully before performing the assay.

Result reliability depends on strict adherence to the instruction for use as described. The following test procedure is only validated for manual procedure. If performing the test on ELISA automatic systems we recommend to perform a whole internal validation process.

8.1. Prior to start the test :

- the distribution and identification plan for all samples should be carefully established on the plate layout supplied in the kit (Annex 1).
- Select the right quantity of strips needed. Let the non used strips in the foil with dessicant for further use.
- Perform all assay steps in the order given and without any delays. Avoid any drying of the wells between steps.
- A clean, disposable tip should be used for dispensing each standard/control and sample.
- Adjust the incubator to 37 ± 1 °C.
- If you are using an automatic plate washer, prime the system. Do not use more than 120 mL of Wash Buffer to prime the washing system.

8.2. Assay procedure :

1. Dispense 100 µL of diluted samples in their respective wells (cf chapter 7.2).
2. Cover the wells with a sticker or cover foil (not provided in the kit).
3. Incubate for 1 hour at 37°C (Plate shaker can be used at 300 rpm).
4. When incubation has been completed, remove the foil, aspirate the content of the wells and wash each well three times with 200 µL of Washing Buffer lavage **WB | CVD**.

Avoid overflows from the reaction wells. The interval between washing and aspiration should be > 5 sec. At the end carefully remove remaining fluid by tapping strips on tissue paper prior to the next step!

Note: Washing is important! Insufficient washing results in poor precision and false results.

5. Dispense 100 µL of diluted conjugate (cf chapter 6.3) in all the wells.
6. Incubate for 1 hour at 37°C. Keep the plate away from the light.
7. Repeat step 4.
8. Dispense 50 µL of Substrate **SU | CVD** in all the wells.
9. Incubate for exactly 15 minutes at room temperature (20-25 °C) in the dark. Blue colour occurs due to an enzymatic reaction.
10. At the end of incubation, remove any remaining fluids by tapping strips upside down on tissue paper. Dispense 200 µL of deionised water and then remove the liquid. Repeat 3 times.
After the 3 cycles, tap the strips on tissue paper.
11. Wait for 15 min at 37°C for drying. Keep the plate away from the light.
12. Read the tests.

8.3. Direct reading of the results

Direct reading of the wells leads to get kind of pictures presented in figure 2.

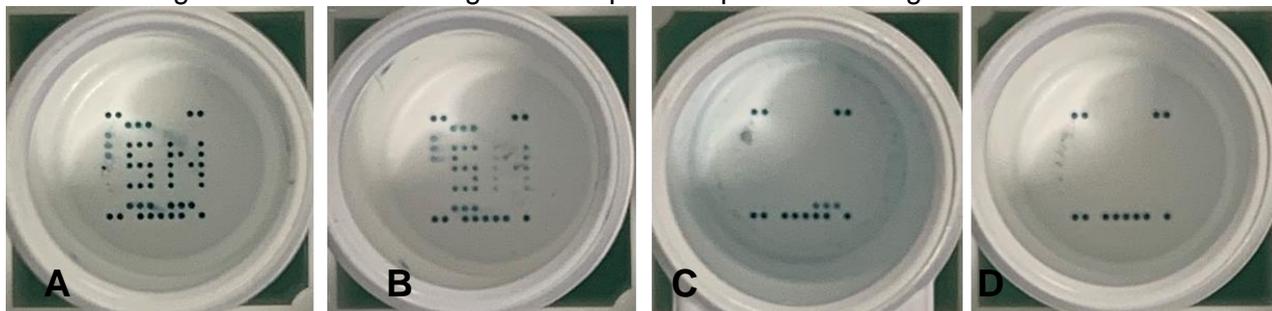


Figure 2 – Examples of pictures of the wells after samples testing.

By using the direct reading sheet (Annex 2), check the boxes when control spots, N and/or S are visible:

- 7 positive controls spots have to be clearly visible. They are validating the testing process. If more than 2 spots are missing, the test can not be validated.
- 5 Sample controls spots (bottom line) have to be clearly visible. They are validating the sample addition. If more than 2 spots are missing, the test can not be validated
- Spots arrayed as a « N » shape are dedicated to the detection of anti-Nucléocapside antibodies. As soon as a « N » is visible, anti-N antibodies are present : Image A, B and C.
- Spots arrayed as a « S » shape are dedicated to the detection of anti-Spike1 antibodies. As soon as a « S » is visible, anti-S1 antibodies are present : Image A and B.

A patient would be seroconverted as soon as N and/or S will be visible.

8.4. Automatic colorimetric reading of the results

The following interpretation process for SARS-CoV-2 IgG Antibody Multiplex ELISA picture analysis is described as example. For our own evaluation studies, 2 colorimetric biochips readers were used:

- SciReader (Scienion, Berlin, Germany)
- SirYus-Reader (Innobiochips, Loos, France).

a- SciReader

We recommend to follow the user guide provided by reader's manufacturer in order to read the tests and extract data. The test interpretation is performed from data provided by the reader.

For the present interpretation, CoViDiag+ 1.0 interpretation template have to be used. In this template, the mean of the net signals from replicates spots for each antigens will be used. Net signals are corresponding to signal measured on each spot and subtracted by background (back ground signal around the antigen spot).

From Mean net signals, define patient status (NEG / BORDER / POS) for each antigen of the test:

| Antigen / (A.U.) | NEG | BORDER | POS |
|-------------------------------------|--------|---------|------|
| Nucléocapside | 0 - 15 | 15 – 30 | > 30 |
| Spike S1 | 0 - 10 | 10 – 20 | > 20 |
| Spike S1 – RBD wt | 0 - 10 | 10 – 20 | > 20 |
| Spike S1 – RBD B.1.1.7 | 0 - 10 | 10 – 20 | > 20 |
| Spike S1 – RBD B.1.351 | 0 - 10 | 10 – 20 | > 20 |
| Spike S1 – RBD B.1.1.248 | 0 - 10 | 10 – 20 | > 20 |
| Spike S1 – NTD | 0 - 10 | 10 – 20 | > 20 |
| Spike S2 | 0 - 15 | 15 – 30 | > 30 |

The cut off values were defined from results obtained by testing 200 plasmas and serums from pre-pandemic healthy subjects (before June 2019).

Interpretation and positivities of samples were done as following :

- **Technical validation of the test** : a test will be validated as soon as the mean of net signals from positive controls: CTRL+ and Samp Ctrl will be above 10 A.U.
- **Sample identified as Positive** :
 - *S1 and/or RBD* and/or NTD is POS*
OR
 - *S2 and/or N is > 40 A.U.*
OR
 - *S2 and/or N is POS and S1 and/or RBD* and/or NTD and/or S2 and/or N is BORDER.*
OR
 - *S2 and/or N is BORDER and S1 and/or RBD* and/or NTD and/or S2 and/or N is BORDER.*

**RBD is defined as POS as soon as one of the RBD (wt or B.1.1.7 or B.1.351 or B.1.1.248) is identified as Positive*

- **Samples identified as Borderline :**
 - *S2 and/or N is POS*
OR
 - *S1 and/or RBD* and/or NTD is BORDER.*
- **Samples identified as Negative :**
All others case not listed above.

b- Lecteur colorimétrique SirYus-Reader

We recommend to follow the user guide provided by reader's manufacturer in order to read the tests and extract data. The test interpretation is performed from data provided by the reader.

For the present interpretation, CoViDiag+ 1.0 interpretation template have to be used. In this template, Raw signals from spots and background (area around the spots) were measured. The mean of the signals and background were calculated for reach antigens and controls. An Index were calculated as below:

$$\text{Mean of raw signals} / (\text{Bckd Mean} + 3 \text{ Bckd Standard Deviation}) \times 100$$

From the Index, NEG/POS status is defined for each antigen or control :

| Spot / Index | NEG | POS |
|-------------------------------------|---------|-------|
| Nucléocapside | 0 - 100 | > 100 |
| Spike S1 | 0 - 100 | > 100 |
| Spike S1 – RBD wt | 0- 100 | > 100 |
| Spike S1 – RBD B.1.1.7 | 0- 100 | > 100 |
| Spike S1 – RBD B.1.351 | 0- 100 | > 100 |
| Spike S1 – RBD B.1.1.248 | 0- 100 | > 100 |
| Spike S1 – NTD | 0 - 100 | > 100 |
| Spike S2 | 0 - 100 | > 100 |

Interpretation and positivities of samples were done as following :

- **Technical validation of the test :** a test will be validated as soon as CTRL+ and Samp Ctrl Index will be > 100.
- **Sample identified as Positive :**
 - *At least 2 Antigens are POS,*
OR
 - *N index is > 150,*
OR
 - *S2 index is > 150.*

**The 4 RBD variants are considered as 1 antigen
RBD is defined as POS as soon as one of the RBD (wt or B.1.1.7 or B.1.351 or B.1.1.248)
index is > 100*

- **Samples identified as Negative :**
All others case not listed above.

9. PRECAUTIONS AND WARNINGS

9.1 Particular attention

- In compliance with article 1 paragraph 2b European directive 98/79/EC the use of the in vitro diagnostic medical devices is intended by the manufacturer to secure suitability, performances and safety of the product. Therefore the test procedure, the information, the precautions and warnings in the instructions for use have to be strictly followed. The use of the test kits with analyzers and similar equipment has to be validated. Any change in design, composition and test procedure as well as for any use in combination with other products not approved by the manufacturer is not authorized; the user himself is responsible for such changes. The manufacturer is not liable for false results and incidents for these reasons. The manufacturer is not liable for any results by visual analysis of the patient samples.
- The present kit is dedicated to in vitro diagnostic use.
- SARS-CoV-2 IgG Antibody Multiplex ELISA is only designed for qualified personnel who are familiar with good laboratory practices.
- All materials of human or animal origin should be regarded and handled as potentially infectious.
- Do not interchange reagents or Microtiterplate strips of different production lots.
- No reagents of other manufacturers should be used along with reagents of this test kit.
- Do not use reagents after expiry dates stated on the label.
- Use only clean pipette tips, dispensers, and lab ware.
- Do not interchange screw caps of reagent vials to avoid cross-contamination.
- Close reagent vials tightly immediately after use to avoid evaporation and microbial contamination.
- After first opening and subsequent storage check conjugate and standard/control vials for microbial contamination prior to further use.
- To avoid cross-contamination and falsely elevated results, pipette patient samples and dispense reagents without splashing into the wells.
- Use only fresh prepared diluted conjugate solution.
- Before any use, check the substrate solution. If the substrate turns blue, it may have been contaminated and should be thrown away.

9.2. Safety note for reagents containing hazardous substances

Reagents may contain <0.05 % of ProClin 300 (See chapter 4.1).
The chromogen substrate of the enzyme is 3,3',5,5'-Tetramethyl-benzidine – TMB.

Therefore, the following hazard and precautionary statements apply.

9.3. Waste disposal

Residues of chemicals and preparations are generally considered as hazardous waste. The disposal of this kind of waste is regulated through national and regional laws and regulations. Contact your local authorities or waste management companies which will give advice on how to dispose hazardous waste.

10. SYMBOLS USED ON THE LABELS

| | |
|---|--|
|  | Storage temperature |
|  | Expiry date |
|  | Read IFU prior to use the kit |
| SC CVD | Precoated plate |
| WB CVD | Wash Buffer |
| DB CVD | Diluant Buffer |
| CA CVD | anti-human IgG Conjugate – HRP (horseradish peroxydase) |
| SU CVD | Chromogenic substrate of the HRP enzyme |
| LOT | Batch Number |

Annexe 1 – Plate layout

| H | G | F | E | D | C | B | A | |
|----------|----------|----------|----------|----------|----------|----------|----------|-----------|
| | | | | | | | | 1 |
| | | | | | | | | 2 |
| | | | | | | | | 3 |
| | | | | | | | | 4 |
| | | | | | | | | 5 |
| | | | | | | | | 6 |
| | | | | | | | | 7 |
| | | | | | | | | 8 |
| | | | | | | | | 9 |
| | | | | | | | | 10 |
| | | | | | | | | 11 |
| | | | | | | | | 12 |

Annexe 2 – Direct reading sheet

Operator name

| | CTRL+ | Ech. | N | S1 |
|------------|-------|------|---|----|
| Wells #A1 | | | | |
| Wells #A2 | | | | |
| Wells #A3 | | | | |
| Wells #A4 | | | | |
| Wells #A5 | | | | |
| Wells #A6 | | | | |
| Wells #A7 | | | | |
| Wells #A8 | | | | |
| Wells #A9 | | | | |
| Wells #A10 | | | | |
| Wells #A11 | | | | |
| Wells #A12 | | | | |
| Wells #B1 | | | | |
| Wells #B2 | | | | |
| Wells #B3 | | | | |
| Wells #B4 | | | | |
| Wells #B5 | | | | |
| Wells #B6 | | | | |
| Wells #B7 | | | | |
| Wells #B8 | | | | |
| Wells #B9 | | | | |
| Wells #B10 | | | | |
| Wells #B11 | | | | |
| Wells #B12 | | | | |
| Wells #C1 | | | | |
| Wells #C2 | | | | |
| Wells #C3 | | | | |
| Wells #C4 | | | | |
| Wells #C5 | | | | |
| Wells #C6 | | | | |
| Wells #C7 | | | | |
| Wells #C8 | | | | |
| Wells #C9 | | | | |
| Wells #C10 | | | | |
| Wells #C11 | | | | |
| Wells #C12 | | | | |
| Wells #D1 | | | | |
| Wells #D2 | | | | |
| Wells #D3 | | | | |
| Wells #D4 | | | | |
| Wells #D5 | | | | |
| Wells #D6 | | | | |
| Wells #D7 | | | | |
| Wells #D8 | | | | |
| Wells #D9 | | | | |
| Wells #D10 | | | | |
| Wells #D11 | | | | |
| Wells #D12 | | | | |

#Plate

| | CTRL+ | Ech. | N | S1 |
|------------|-------|------|---|----|
| Wells #E1 | | | | |
| Wells #E2 | | | | |
| Wells #E3 | | | | |
| Wells #E4 | | | | |
| Wells #E5 | | | | |
| Wells #E6 | | | | |
| Wells #E7 | | | | |
| Wells #E8 | | | | |
| Wells #E9 | | | | |
| Wells #E10 | | | | |
| Wells #E11 | | | | |
| Wells #E12 | | | | |
| Wells #F1 | | | | |
| Wells #F2 | | | | |
| Wells #F3 | | | | |
| Wells #F4 | | | | |
| Wells #F5 | | | | |
| Wells #F6 | | | | |
| Wells #F7 | | | | |
| Wells #F8 | | | | |
| Wells #F9 | | | | |
| Wells #F10 | | | | |
| Wells #F11 | | | | |
| Wells #F12 | | | | |
| Wells #G1 | | | | |
| Wells #G2 | | | | |
| Wells #G3 | | | | |
| Wells #G4 | | | | |
| Wells #G5 | | | | |
| Wells #G6 | | | | |
| Wells #G7 | | | | |
| Wells #G8 | | | | |
| Wells #G9 | | | | |
| Wells #G10 | | | | |
| Wells #G11 | | | | |
| Wells #G12 | | | | |
| Wells #H1 | | | | |
| Wells #H2 | | | | |
| Wells #H3 | | | | |
| Wells #H4 | | | | |
| Wells #H5 | | | | |
| Wells #H6 | | | | |
| Wells #H7 | | | | |
| Wells #H8 | | | | |
| Wells #H9 | | | | |
| Wells #H10 | | | | |
| Wells #H11 | | | | |
| Wells #H12 | | | | |